

# Cinnamoyl Derivatives from *Cordia Platythyrsa* and Chemiotaxonomical Value of the *Cordia* Genus

Bernard Dabole<sup>1,\*</sup>, Rostanie Zeukang<sup>2</sup>, Alex de Theodore Atchade<sup>2</sup>, Turibio Tabopda<sup>2</sup>, Benoit Bargui Koubala<sup>1</sup>, Joseph Tanyi Mbafor<sup>2</sup>

<sup>1</sup>Department of Chemistry, University of Maroua, Maroua, Cameroon

<sup>2</sup>Department of Organic Chemistry, University of Yaounde-I, Yaounde, Cameroon

## Email address:

dabolebernard@yahoo.fr (D. Bernard)

\*Corresponding author

## To cite this article:

Bernard Dabole, Rostanie Zeukang, Alex de Theodore Atchade, Turibio Tabopda, Benoit Bargui Koubala, Joseph Tanyi Mbafor. Cinnamoyl Derivatives from *Cordia Platythyrsa* and Chemiotaxonomical Value of the *Cordia* Genus. *Science Journal of Chemistry*.

Vol. 4, No. 3, 2016, pp. 36-40. doi: 10.11648/j.sjc.20160403.12

Received: June 6, 2016; Accepted: June 15, 2016; Published: June 30, 2016

**Abstract:** Phytochemical investigation of the roots and stem barks of *Cordia platythyrsa* (Boraginaceae) had led to the isolation of two new cinnamates, the cordicinnamate **A** compound 1 and the cordicinnamate **B** compound 2 along with four known compounds. Their structures were established by spectroscopic analysis mainly FAB and TOF –MS, <sup>1</sup>H NMR and <sup>13</sup>C NMR, COSY, HMBC, HSQC and by comparison with literature data. The cinnamoyl derivatives were reported for the first time in the *Cordia* genus. The isolation and identification of cinnamoyl derivatives in *cordia* genus improve the chemiotaxonomy value in this genus.

**Keywords:** Boraginaceae, *Cordia Platythyrsa*, Cinnamoyl Derivatives, Chemiotaxonomy

## 1. Introduction

The genus *cordia* belongs to the family of boraginaceae and it is found in warm regions [1]. The decoction of several species of *cordia* genus has been used in traditional medicine to treat influenza, fever, pneumonia, coughs, insomnia, stomach-ache, parasitic [2] and infections [3]. Previous phytochemical investigations of *cordia* genus led to the isolation of pyrrolizidine alkaloids, terpenoids flavonoids, lignans, meroterpenoids naphthoquinones [4]. In addition, phytochemical studies of *Cordia platythyrsa* have revealed the presence of shingolipids, Cordiachromes A-F [5] and xanthones [6]. Its leaves are used for the treatment of convulsions and sleeping sickness (maceration) [7]. With a view to extending the phytochemical investigations of these species, the present paper report the isolation and structure characterization of two new cinnamates namely cordicinnamate A-B and other known compounds.

## 2. Experimental

### 2.1. General Experimental Procedure

Melting points were determined on a Büchi 434 melting point apparatus and were uncorrected. <sup>1</sup>H NMR (400 MHz) and <sup>13</sup>C NMR (150MHz) spectra were recorded at room temperature in CDCl<sub>3</sub> using a Bruker AVANCE AM 400 and AMX 500 NMR instruments. Chemical shifts are given in  $\delta$  (ppm) value relative to TMS as internal standard. ESI-TOF mass spectra operating in positive mode were recorded on a finnigan MAT 312 and FAB mass spectra was recorded on Jeol JMS HX 110 mass spectrometer. Silica gel (230-400 meshes) and sephadex gel (LH-20) were used for Column Chromatography (CC) and vacuum liquid chromatography (VLC). Thin Layer Chromatography (TLC) was performed on precoated silica gel plates (60 F254, Macherey-Nagel) using the system solvent n-hexane – EtOAc (9.2:0.8, 8:2) and EtOAc-MeOH (6.5:3.5) as eluent. Spots were visualized by UV light ( $\lambda_{max}$  = 254 nm, 366 nm) and were observed after using sulphuric acid (50%) as spraying reagent.

## 2.2. Plant Material

The stem barks and roots of *Cordia platythyrsa* were collected in Yaoundé town (Cameroon) and were identified by Dr N. Tsabang, of the Centre for Study of Medicinal Plants of Yaoundé (Cameroon). One voucher specimen (N<sup>o</sup>43625/HNC) was deposited at the National Herbarium of Cameroon (NHC).

## 2.3. Extraction and Isolation

The powdered roots (2 Kg) were extracted with CH<sub>2</sub>Cl<sub>2</sub>-MeOH (1:1) at room temperature for 72 hours. The resulting extract was evaporated to dryness under reduced pressure to yield a dark residue (12.72 g). This residue was partitioned with n-hexane, EtOAc and MeOH. The EtOAc extract was evaporated under reduced pressure to afford 8 g of residue. This residue was submitted to vacuum liquid chromatography (VLC) on silica gel 230-400 meshes eluted with n-hexane – EtOAc and EtOAc in order of increasing polarity. As result, 3 fractions (S1, S2 and S3) were collected on the basis of TLC. Fraction S3 was subjected to CC using silica gel 230-400 meshes eluted with n-hexane – EtOAc (9.2:0.8) afforded to 9 subfractions. Subfraction S375 was purified using sephadex gel LH-20 eluted with MeOH to afford compounds 1 (1.90 mg) and 5 (2 mg).

The powdered stem barks (6.5 Kg) were extracted with CH<sub>2</sub>Cl<sub>2</sub>-MeOH (1:1) at room temperature for 72 hours. The resulting extract was evaporated to dryness under reduced pressure to yield a dark residue (200 g). This residue was partitioned with n-hexane, EtOAc and MeOH. The EtOAc extract was evaporated under reduced pressure to give 97 g of residue which was in turn submitted to vacuum liquid chromatography (VLC) on using silica gel 230-400 meshes eluted with n-hexane – EtOAc, EtOAc and EtOAc- MeOH in order of increasing polarity. As result, 7 fractions (B0, B1, B2, B3, B4, B5 and B6) were collected on the basis of TLC. Fraction B3 and B4 were subjected to CC purification using silica gel 230-400 meshes eluted with n-hexane – EtOAc (8:2) to give compounds 2 (2mg), 3 (2mg) and 6 (3mg). Fraction B6 was subjected to CC purification using silica gel 230-400 meshes eluted with EtOAc- MeOH (6.5:3.5) afforded to 4 (20mg).

## 3. Results and Discussion

### 3.1. Identification of the Compounds

Compound 1 was obtained as a white powder. Its molecular formula was deduced as C<sub>28</sub>H<sub>46</sub>O<sub>3</sub> by negative mode FAB-MS at m/z = 429.0 [M-H]<sup>+</sup>. The cinnamoyl moiety was characterized by <sup>1</sup>HNMR signals (table 1) at δ<sub>H</sub> 6.81 (1H,d, J=8Hz, H-2'), 6.99 (1H, d, J=8Hz, H-3'), 6.97(1H,S, H-5'), 6.21(1H,d, J=16Hz, H-2) and 7.53 (1H, d, J=16Hz, H-3) [8, 9]. The methoxy and methyl group linked to aromatic ring were observed respectively at δ<sub>H</sub> 3.85 (3H,

S, 4'-OCH<sub>3</sub>) and 1.53 (3H, S, 6'-CH<sub>3</sub>). Proton signals owing to aliphatic chain were observed at δ<sub>H</sub> 1.18(12H, S, H-5''-14''), 4.11(2H, t, H-1'') and 0.81(6H, S, 2x4''-CH<sub>3</sub>). The <sup>13</sup>C NMR spectrum with signals at δ<sub>C</sub> 168,0 (C, C-1), 114,7(CH,C-2'), 123,1 (CH,C-3'), 109,2 (CH,C-5'), δ 115,7 (CH,C-2) and 144,0 (CH, C-3) had demonstrated the presence of cinnamoyl moiety [8, 9]. Aliphatic chain was identified and characterized by signals at δ 22.7, 31.9, 23.0 (10 CH<sub>2</sub>, C-5''-14''), 37.2 (C, C-4''). Protons position of aliphatic chain and cinnamoyl moiety were supported by COSY correlations (figure 1) H-2'/ H-3, H-2'/H-3' and H-1''/H-2''. The carbon signals were assigned by HSQC spectrum and were confirmed by HMBC spectrum. Using HMBC correlations (figure 1) of H-3''/C-4''; 4'-OCH<sub>3</sub>/C-4'; 6'-CH<sub>3</sub>/C-6' and H-1''/C-1, C-2'' had permitted to link aliphatic chain, methoxy and methyl group to cinnamoyl moiety. From the spectral evidences and literature data, the structure of compound 1 was characterized as 1-(4'',4''-dimethylpentadecyl)-4'-methoxy-6'-methyl cinnamate (Cordicinnamate A).

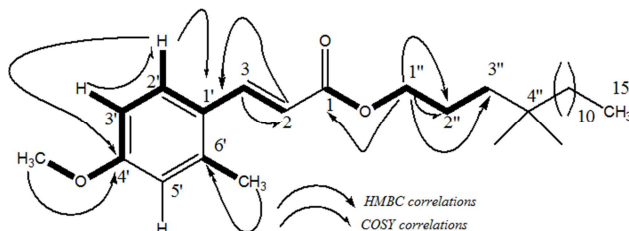


Figure 1. HMBC and COSY correlations (bold lines) for compound 1.

Compound 2 isolated as a white powder, exhibited molecular formula C<sub>39</sub>H<sub>68</sub>O<sub>4</sub> as determined from its positive mode TOF-MS at m/z = 603.5003 [M+3H]<sup>+</sup>. Comparison of the NMR data (table 1) of compound 2 with those of compound 1 indicated a slight modification in the cinnamoyl moiety of compound 2. Instead of the aromatic proton at C-3' and methyl group at C-6', signals detected were those of an aliphatic chain and aromatic proton at δ<sub>H</sub> 1.64 (2H, m, H-1'''), 1.18 (36H, S, H-3'''-21'''), 1.53 (2H, m, H-22'''), 3.53 (2H, t, H-23''') and 7.03(1H, d, J=8 Hz, 1.5 Hz, d, H-6') respectively in <sup>1</sup>HNMR spectrum. HSQC NMR experiment was used to assign H-1''', H-3'''-21''', H-22''', H-23''' and H-6'. <sup>1</sup>HNMR spectrum showed signals at δ<sub>H</sub> 3.32 (3H, S, 23'''-OCH<sub>3</sub>) characterize methoxy group attached to aliphatic chain and at δ<sub>H</sub> 1.48 (2H, m, H-2'') and 1.32 (2H, m, H-4'') attributed to a second aliphatic chain. Protons positions of the above aliphatic chain and cinnamoyl moiety were confirmed by COSY correlations (figure 2) H-2'/H-3, H-5'/H-6', H-1''/H-2'' and H-22'''/H-23'''. The HMBC correlation (figure 2) of H-5'/C-1', C-3 and H-1''/C-2'', C-4'' demonstrated the presence of ester group [8, 9]. Thus the structure of 2 was established as 3'-(23'''-methoxy tricicosanyl)-1-(pentyl)-4'-methoxycinnamate (Cordicinnamate B).

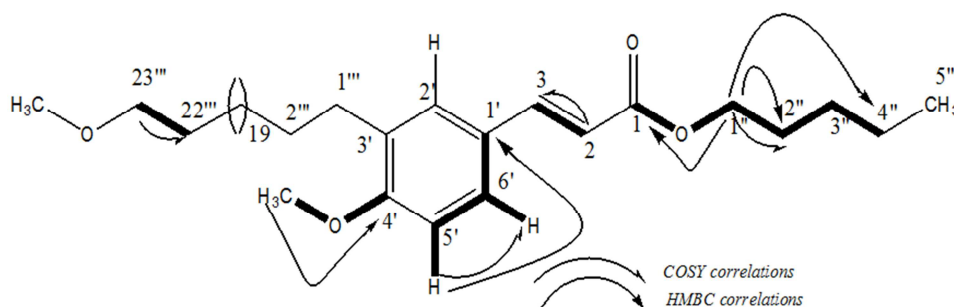


Figure 2. HMBC and COSY correlations (bold lines) for compound 2.

Compound 3 was obtained as a white powder. Its molecular formula was assigned as  $C_{15}H_{20}O_5$  as deduced from TOF-MS (positive mode)  $m/z = 282$   $[M+2H]^+$ . Comparison of the NMR data (table 1) of compound 3 with those of compound 2 indicated the signal at  $\delta_H$  3.91 (1H, s, 3'-OCH<sub>3</sub>), 5.81 (1H, s, 4'-OH) and 3.61 (2H, t) instead of aliphatic chain, methoxy and methylene group respectively in  $^1H$ NMR spectrum. Its structure (Figure 3) was elucidated by 1D and 2D NMR spectroscopy and it was identified as 1-(5''-hydroxypentyl)-4-hydroxy-3-methoxycinnamate [10]. Structures of the compounds 4-6 were elucidated by 1D and 2D NMR spectroscopy. All the physical and spectral data were identical to the reported values in literature [6, 11-14]. The compounds were identified as methyl orsellinate 4, paramethoxytoylehexanoate 5 and 3-O- $\beta$ -D-glucopyranosyl- $\beta$ -sitosterol 6 (Figure 3).

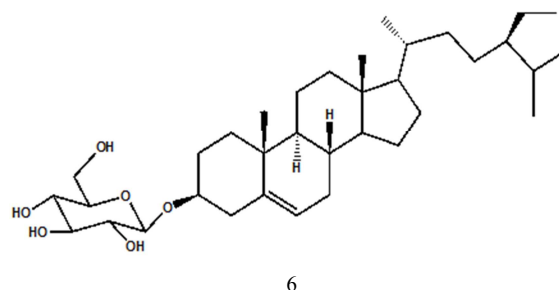
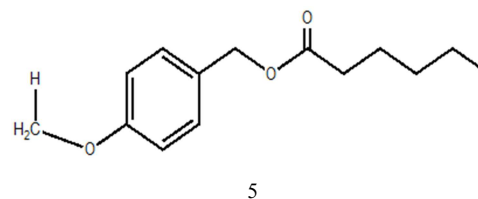
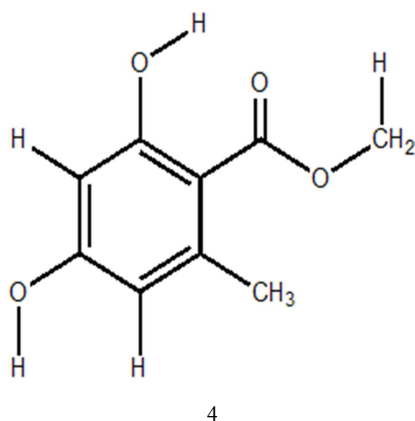
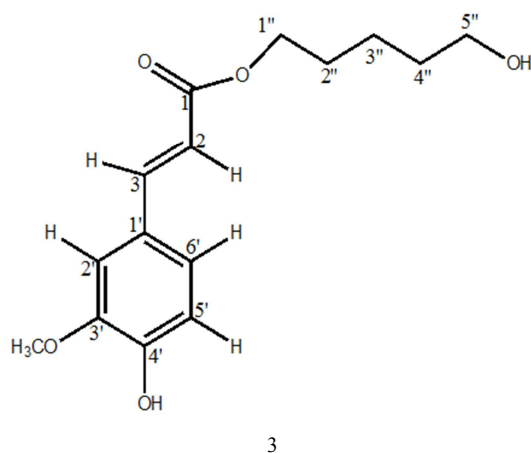


Figure 3. Chemical structure of compound 3-6.



Previous studies of *Cordia* species led to various skeletons of compounds. The triterpenoids have been isolated both from *Cordia verbenacea* DC and *Cordia multispicata* [15] and Dammarane-type triterpenes both from *Cordia spinescens* and *Cordia multispicata* [16]. The Meroterpenoid naphthoquinones have been isolated both from the roots of *Cordia linnaei* and *Cordia corymbosa* [17]. The roots of *Cordia curassavica* [18] and wood of *Cordia fragrantissima* afforded hydroquinones [19]. The magnesium lithospermate, Calcium rosmarinate and magnesium rosmarinate were isolated from the leaves of *Cordia spinescens* [20]. The investigation of the heartwood of *Cordia millenii* yielded terpenoid quinones [21]. Hydroquinone terpenoids were isolated both from *Cordia alliodora* and the heartwood of *Cordia elaeagnoides* [22]. The terpenoid glycosides have been isolated from *Cordia oblique* [23].

In addition, phytochemical studies on *Cordia platythyrsa* yielded to two new cinnamoyl derivatives, Cordicinnamate A (compound 1) and Cordicinnamate B (compound 2). To the best of our knowledge this is the first time that cinnamates have been reported from *Cordia* genus.

### 3.2. Cordicinnamate A (Compound 1)

White powder; MP = 116-118°C;  $^1H$  NMR ( $CDCl_3$ , 400MHz) and  $^{13}C$  NMR ( $CDCl_3$ , 150MHz) data see table 1; FAB-MS  $m/z = 429.0$   $[M-H]^+$  (calcd. for  $C_{28}H_{46}O_3$ , 430.0).

**Table 1.**  $^1\text{H}$  NMR (400MHz) and  $^{13}\text{C}$  NMR (150MHz) data for compounds 1-3 in  $\text{CDCl}_3$ .

| 1                      |                  |                           | 2          |                            |            | 3                          |  |  |
|------------------------|------------------|---------------------------|------------|----------------------------|------------|----------------------------|--|--|
| Position               | $\delta_C$       | $\delta_H$ [mult, J (Hz)] | $\delta_C$ | $\delta_H$ [mult, J (Hz)]  | $\delta_C$ | $\delta_H$ [mult, J (Hz)]  |  |  |
| 1                      | 168.0            | -                         | 168.0      | -                          | 167.0      | -                          |  |  |
| 2                      | 115.7            | 6.21 (1H, d, J =16Hz)     | 115.0      | 6.21(1H, d, J =16Hz)       | 117.0      | 6.27(1H, d, J =15.5Hz)     |  |  |
| 3                      | 144.0            | 7.53(1H, d, J =16Hz)      | 144.0      | 7.57(1H, d, J =16Hz)       | 144.0      | 7.58(1H, d, J =15.5Hz)     |  |  |
| 1'                     | 127.0            | -                         | 127        | -                          | 127.0      | -                          |  |  |
| 2'                     | 114.7            | 6.81(1H, d, J =8Hz)       | 109.0      | 7.00(1H, d, J =1.5Hz)      | 109.0      | 7.01(1H, d, J =1.6Hz)      |  |  |
| 3                      | 123.1            | 6.99(1H, d, J =8Hz)       | 148        | -                          | 146.0      | -                          |  |  |
| 4'                     | 147.0            | -                         | 147.0      | -                          | 115.0      | -                          |  |  |
| 5'                     | 109.2            | 6.97(1H, S, 1H)           | 114.0      | 6.76(1H, d, J =8Hz)        | 123.0      | 7.05(1H, d, J =8Hz)        |  |  |
| 6'                     | 135.0            | -                         | 122.5      | 7.03(1H, d, J =1.5Hz, 8Hz) | 116.0      | 6.86(1H, d, J =1.6Hz; 8Hz) |  |  |
| 3'-OCH <sub>3</sub>    | -                | -                         | -          | -                          | 56.0       | 3.91(1H, S)                |  |  |
| 4'-OH                  | -                | -                         | -          | -                          | -          | 5.81(3H, S)                |  |  |
| 4'-OCH <sub>3</sub>    | 55.9             | 3.85(3H, S)               | 56.0       | 3.85(3H, S)                | -          | -                          |  |  |
| 6'-CH <sub>3</sub>     | 28.8             | 1.53(3H, S)               | -          | -                          | -          | -                          |  |  |
| 1''                    | 64.6             | 4.11(1H, t)               | 64.6       | 4.11(2H, t)                | 64.5       | 4.16(2H, t)                |  |  |
| 2''                    | 26.0             | 1.60(2H, m)               | 29.0       | 1.48(2H, m)                | 30.0       | 1.6(2H, m)                 |  |  |
| 3''                    | 28.0             | 0.86(2H, m)               | 23.0       | 1.28(2H, m)                | 26.0       | 1.2-1.5(2H, m)             |  |  |
| 4''                    | 37.2             | -                         | 26.0       | 1.32(2H, m)                | 25.9       | 1.5(2H, m)                 |  |  |
| 5''                    | -                | -                         | 14.0       | 0.81(3H, t)                | 64.0       | 3.61(2H, m)                |  |  |
| 5''-14''               | 22.7, 23.0, 31.9 | 0.71-0.86(12H,m)          | -          | -                          | -          | -                          |  |  |
| 15''                   | 14.2             | 0.78(3H, m)               | -          | -                          | -          | -                          |  |  |
| 4''-CH <sub>3</sub>    | 19.4             | 0.75(3H, m)               | -          | -                          | -          | -                          |  |  |
| 4''-CH <sub>3</sub>    | 18.8             | 0.75(3H, m)               | -          | -                          | -          | -                          |  |  |
| 1'''                   | -                | -                         | 34.5       | 2.20(2H, t)                | -          | -                          |  |  |
| 2'''                   | -                | -                         | 28.0       | 1.64(2H, m)                | -          | -                          |  |  |
| 3'''-21'''             | -                | -                         | 23.0       | 1.18(36H, S)               | -          | -                          |  |  |
| 22'''                  | -                | -                         | 25.0       | 1.53(2H, m)                | -          | -                          |  |  |
| 23'''                  | -                | -                         | 62.5       | 3.53(2H, t)                | -          | -                          |  |  |
| 23'''-OCH <sub>3</sub> | -                | -                         | 49.5       | 3.32(3H, S)                | -          | -                          |  |  |

### 3.3. Cordicinnamate B (Compound 2)

White powder; MP =185°C;  $^1\text{H}$  NMR ( $\text{CDCl}_3$ , 400MHz) and  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 150MHz) data see table 1; TOF-MS  $m/z$  = 603.5003  $[\text{M}+3\text{H}]^+$  (calcd. for  $\text{C}_{39}\text{H}_{68}\text{O}_4$ , 600.5003).

### 3.4. 1-(5''-hydroxypentyl)-4-hydroxy-3-methoxy Cinnamate (Compound 3)

White powder; MP = 63-64°C;  $^1\text{H}$  NMR ( $\text{CDCl}_3$ , 400MHz) and  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 150MHz) data see table 1; TOF-MS  $m/z$  = 282  $[\text{M}+2\text{H}]^+$  (calcd. for  $\text{C}_{15}\text{H}_{20}\text{O}_5$ , 280.0).

## 4. Conclusion

Phytochemical studies on *Cordia platythyrsa* yielded two cinnamoyl derivatives isolated for the first time on *Cordia* genus along with five others known compounds. To the best of our knowledge, this is the first time to isolate the cinnamoyl skeleton on the *Cordia* genus. The isolation and the identification of cinnamoyl derivatives in *Cordia* genus improve the chemiotaxonomy value of the *Cordia* genus. Further studies aiming to isolate and identify compounds for

the other fractions are needed.

## Acknowledgments

The authors would like to thank Prof. Dr. Muhammad Iqbal Choudhary, the TWAS (the Academy of Sciences for the Developing World), the H. E. J (Research Institute of Chemistry), ICCBS (Center for Chemical and Biological Sciences), the University of Karachi, Karachi-75270, Pakistan, the University of Yaoundé-I, Cameroon and the University of Maroua, Cameroon; for the fellowships we receive from them and their notable contribution on these studies.

## References

- [1] Thirupathi, K., Kumar S., Raju V., Ravikumar B., Krishna D., Mohan G.. A review of medicinal plants of the genus *Cordia*: their chemistry and pharmacological uses. *J. Nat. Rem.*, 8, 2008, pp. 1-10.
- [2] Dalziel, J., Hutchinson, J. The useful plants of west tropical Africa an appendix to the flora of West Africa, 1948, pp 424.

- [3] Al-Awadi, F., Srikumar, T., Anim, J., Khan, I. Antiinflammatory Effects of *Cordia myxa* Fruit on Experimentally Induced Colitis in Rats. *Nutrition* 17, 2001, pp. 391–396.
- [4] Okusa, P., Penge O., Duez, P., Devleeschouwer, M. Direct and indirect antimicrobial effects and antioxidant activity of *Cordia gillettii* De Wild (Boraginaceae). *J. Ethnopharm* 112, 2007, pp. 476–481.
- [5] Tapondjou, L., Mitaine-offer, A., Sautor M., Tomofumi M., Lacaille-Dubois A. Shingolipids and other constituents from *Cordia platythyrsa*. *Biochem. Syst. and Ecol.* 33, 2005, pp. 1293–1297.
- [6] Atchadé, T., Dabolé B., Achyut, A., Kahn, A., Mbafor J., Choudhary M. Chemical Constituents of *Cordia Platythyrsa* and evaluation of their glycation and urease inhibition activities. *J. Nat. Prod. Ind.*; 8, 2012, pp. 346–351.
- [7] Hutchinson J., Dalziel J. M., «Flora of West Tropical Africa», Mill Bank, Londre, 1963, pp. 317–32.
- [8] Somepalli V., Merallapudi S., Alluri V., Golakoti T., Gottumukkala V. Antioxidant and antimicrobial activity evaluation of polyhydroxycinnamic acid ester derivatives, *Ind. J. Chem.* 45B, 2005, pp. 252–257.
- [9] Talla Emmanuel, DaoudaDjibo., Nyemb Jean-Noël, Sophie Laurent, Vander Eist Luce, Dabole Bernard and MbaforTanyi Joseph. Two new compounds from stem barks of *Vepris heterophylla* (Engl.) R. Let. (Rutaceae). *Journal of Chemical and Pharmaceutical Research*, 7(7), 2015, pp. 553–557.
- [10] Ronchetti D., Impagnatiello F., Guzzetta M., Gasparini L., Borgatti M., Gambari R., Ongni E.. Modulation of INOS expression by a nitric oxide-releasing derivative of the natural antioxidant ferulic acid in activated RAW 264.7 macrophages. *European J. Pharm* 12, 532, 2006, pp. 162–169.
- [11] Faizi S., Ali M., Saleem R., Irfanullah, Bibi S., «Complete <sup>1</sup>H and <sup>13</sup>C NMR assignments of stigma-5-en-3-O-β-glucoside and its acetyl derivative», *Magnetic Resonance Chemistry*, 39, 2001, pp.399–405.
- [12] Ingólfssdóttir K., Gudmundsdóttir G. F., Ögmundsdóttir H. M., Paulus K., Haraldsdóttir S., Kristinsson H., and Bauer R., «Effects of teuorin and methyl orsellinate from the lichen *Peltigera leucophlebia* on 5-/15-lipoxygenases and proliferation of malignant cell lines in vitro». *Phytomedicine*, 9, 2002, pp. 654–658.
- [13] Rojas I. S., Lotina-Hennsen B., Mata R., «Effect of licheb metabolites on thylakoid electron transport and photophosphorylation in isolated spinach chloroplasts 1». *Journal of Natural Products*, 63, (2000), 1396–1399.
- [14] Sandjo Louis Pergaud, «Sphingolipides, Triterpénod's et autres métabolites secondaires des variétés sauvage et cultivée de l'espèce *Triumfetta cordifolia* A. Rich. (Tiliaceae): Transformations chimiques et évaluation des propriétés biologiques de quelques composés isolés», Thèse de Doctorat /Ph.D en Chimie Organique, Université de Yaoundé I-Cameroun, 2009, pp. 33.
- [15] Masanori Kuroyanagi, Takahiro Seki, Tatsuo Hayashi, Yoshio Nagashima, Nobuo Kawahara, Setsuko Sekita, and Motoyoshi Satake. «Anti-androgenic Triterpenoids from the Brazilian medicinal plant *Cordia multispicata*». *Chem. Pharm. Bull.*, 49(8), 2001, pp. 954–957.
- [16] Masanori Kuroyanagi, Nobuo Kawahara, Setsuko Sekita, Motoyoshi Satake, Tatsuo Hayashi, Yoichi Takase, and Kazuo Masuda, «Dammarane-type triterpenes from *Cordia Spinescens*». *Journal of Natural Products*, 66, 2003, pp. 1307–1312.
- [17] Lothar W. Bieber, Hans C. Krebs and Wolfram Schäfer, «Further Meroterpenoid naphthoquinones from *cordia corymbosa*». *Phytochemistry*, 35(4), 1994, pp. 1027–1028.
- [18] Ioset J. R., Marston A., Gupta M. P., Hostettmann K., «Antifungal and larvicidal cordiaquinones from the roots of *Cordia curassavica*». *Phytochemistry*, 53, 2000, pp. 613–6
- [19] Mori K., Kawano M., Fuchino H., Ooi T., Satake M., Agatsuma Y., Kusumi T., and Sekita S. «Antileishmanial Compounds from *Cordia fragrantissima* collected in Burma (Myanmar) ». *Journal of Natural Product*, 71, 2008, pp. 18–21.
- [20] Yasmina Aura Lim, Shiho Kojima, Norio Nakamura, Hirotosugu Miyashiro, Hirotoshi Fushimi, Katsuko Komatsu, Masao Hattori, Kunitada Shimotohno, Mahabir P. Gupta and Mireya Correa, « Inhibitory effects of *Cordia spinescens* extracts and their constituents on reverse transcriptase and protease from human immunodeficiency virus». *Phytotherapy research*, 11, 1997, pp. 490–495.
- [21] Moir M. and Ronald H. Thomson, «Naturally occurring quinines. Part XXII. Terpenoid quinones in *Cordia* Spp.». *Journal of Chemical Society Perkin I.*, 1973, pp. 1352–1357.
- [22] Gary D. Manners and Leonard Jurd, «The Hydroquinone Terpenoids of *Cordia alliodora*». *Journal of Chemical Society Perkin Transcent I*, 1997, pp. 405–410.
- [23] Srivastava S. K., Srivastava S. D. and Nigam S. S., Lupa-20(29)-ene-3-O-α-L-rhamnopyranoside from the roots of *Cordia oblique*. *J. Indian Chem. Soc.*, LX, 1983, pp. 202.